Original Research Article

Leptospira culture and isolation from tertiary care hospital, Surat

Alpa S Patel^{1,*}, Summaiya A Mulla², Hardik C Patel³¹Dept. of Microbiology, Government Medical College, Surat, Gujarat, India²Dept. of Microbiology, Government Medical College, Jamnagar, Gujarat, India³S.R.B Shah Mahavir Hospital, Surat, Gujarat, India

ARTICLE INFO

Article history:

Received 22-02-2019

Accepted 05-02-2021

Available online 02-04-2021

Keywords:

Leptospirosis

Isolation

EMJH medium

Blood culture

ABSTRACT

Background: In a highly globalized world, where boundaries are continually crossed due to different international activities, tropical diseases including Leptospirosis have emerged as significant causes of severe illness and death in untreated patients.

Aim: To optimize the culture isolation of Leptospira by using different culture methods.

Materials and Methods: All suspected cases of Leptospirosis regardless of their age/sex were recruited following informed and written consent. Five different blood culture methods have been tried to see better isolation of Leptospira from patient.

Result: Total 326 patients are evaluated over a period of three years. Five different method of blood culture were followed out of that, whole blood culture technique shows better result compare to other followed technique.

Conclusion: The culture methods are gold standard but time consuming. It has an important role in the study of outbreaks. It provides epidemiological data that is helpful in studying pathogenesis of Leptospira.

© This is an open access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>) which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

1. Introduction

Leptospirosis is a bacterial zoonotic disease. It is increasingly recognized as emerging or re-emerging disease. Particularly common in rural tropical regions associated with exposure to contaminated water and soil during heavy rainfall and occupational risks. Because of its multiple and non-specific clinical presentation, Medical diagnosis of Leptospira is especially challenging and clinical suspicions warrant systematic biological confirmation.^{1–3} The clinical manifestations of leptospirosis range from a mild influenza-like illness to multi-organ failure and death. The common signs and symptoms of this infection is difficult to discriminate leptospirosis from a range of other infectious diseases that occur in the tropics, including dengue fever, malaria, and rickettsial infection.⁴

In acute stage, Leptospira isolation from clinical samples has been considered the gold standard for decades. However, leptospira are fastidious and slow-growing organisms. The cultures are to be kept and Screened for up to 14 weeks,⁵ resulting in low isolation sensitivity,⁴ so that isolation is of limited use for medical diagnosis and treatment. There is a great value of gaining access to bacterial isolates: they provide biological material for epidemiology (serological and/or molecular) as well as research for purpose. (e.g. virulence, comparative genomics).

2. Materials and Methods

Leptospirosis is a zoonotic disease of worldwide distribution. Leptospirosis is primarily a contagious disease of animals, accidental infection occurs to humans and is caused by pathogenic spirochete of the genus leptospira that traditionally consist of two species *L.interrogans* and

* Corresponding author.

E-mail address: dralpain@gmail.com (A. S. Patel).

L.biflexa. *L.interrogans* includes all pathogenic serovars and *L.biflexa* includes all the saprophytic strains.⁶ Each sub-group consists of several strains designated as Serovars. The predominant serovars of leptospira present in India are *L.andamana*, *L.pomona*, *L.grippityphosa*, *L.hebdomadis*, *L.semoranga*, *L.javanica*, *L.autumnalis*, *L.canicola*.⁷

Leptospirosis is usually a seasonal disease that starts at the onset of the rainy season, the disease declines as the rains recede. Infection is acquired through direct contact with the environment contaminated with urine of rodents, carrier or diseased animals. Agricultural workers such as rice field planters, sugar cane and pineapple field harvesters, livestock handlers, laborers engaged in canal cleaning operations are subjected to exposure with leptospira.⁶ Males suffer more frequently from leptospirosis than females because of more out-door activity and greater occupational exposure.

We have conducted a prospective study from September 2011 to October 2013, at New civil hospital, Government Medical college, Surat. The study protocol was approved by the Human Research Ethics Committee of the college. All suspected cases of Leptospirosis regardless of their age/sex were recruited following informed and written consent. A detailed history and the physical examination findings were recorded on a clinical record form.

2.1. Sample collection for blood culture

The anti-cubital vein of either hand, will first be cleaned with 70% alcohol in concentric manner from inside out and allowed to dry. Then tincture iodine will be applied in same manner. Again alcohol will be applied and let it dry. Approximately 2ml of patient's blood will be collected. Preferably sample for blood culture will be collected before administration of antibiotics.

2.2. Procedure to be followed for sample collection for blood culture

After collecting blood with all aseptic precautions. Drop of whole blood is directly inoculated into 4ml liquid EMJH medium (Ellinghausen, McCullough, Johnson, Harris) between flames of two spirit lamp bed side to decrease the contamination. While 1ml of sample is collected in Heparin sodium vacuette containing 250 µl of heparin. Another 1ml of sample is collected in plain vacuette.

2.3. Four different sampling methods were used to culture from heparinevacuette⁸

1. Drop of heparin whole blood will be placed directly into 4ml liquid EMJH medium (will be known as 1st number tube of heparin whole blood).
2. Remaining 1900 µl blood will be centrifuge at 3,000 rpm for 10 min & left undisturbed for 30 min, 10 drops of surface plasma will be added to 4 ml of liquid EMJH medium (will be known as 1st number tube of surface

plasma).

3. Remaining 1400µl of plasma centrifuge at 6000 rpm for 3min, supernatant will be removed and 4 drops of deposit will be added into 4 ml of liquid EMJH medium (will be known as 1st number tube of spun plasma).
4. For clot culture, the serum from plain vacuette will be separated by sterile tips, now the clott will be break by sterile glass beads and transferred to 4ml of liquid EMJH (will be known as 1st number tube of clot culture).
 - a. 1st Subculture will be done within 6hrs of collection from 1st number tube of each set, in 4ml of liquid EMJH and it will be numbered as tube number 2, taking 1ml of inoculum by sterile syringe & needle of 2ml. Simultaneously screening will also be done from the remaining culture of the same syringe and needle.
 - b. 2nd Subculture will be done on next day from 2nd number tube, in 4ml of semi-solid EMJH medium and will be numbered as tube no 3, taking 1ml of inoculum by sterile syringe & needle of 2ml.
 - c. 3rd Subculture will be done on 3rd day from 3rd number of tube and will be numbered as 4, in 4ml of semi-solid EMJH medium, taking 1ml of inoculum by sterile syringe & needle of 2ml.
 - d. 4th Subculture will be done from 4th number of tube on 5th day and will be numbered as 5th tube, in 4ml of semi-solid EMJH medium, taking 1ml of inoculum by sterile syringe & needle of 2ml.
 - e. Figure 2 Liquid and Semi – solid EMJH medium.

All cultures will be incubated aerobically and protected from light at room temperature (28 to 30°C) for six months; the cultures will be examined weekly for three months and then every 2 to 4 weeks for a further 3 months by placing one drop of the culture onto a microscopic glass slide and observing it by dark-field microscopy. If any contamination will be seen macroscopically or microscopically cultures will be filtered with 0.22µm filters. The flow chart no – 1 shows, processing of blood culture sample.

2.4. Follow up of sample

1. The cultures are examined weekly for 3 months and then every 2 to 4 weeks for 3 months by placing one drop of the culture onto a microscopic slide and observing it by dark-field microscopy at ×200 magnification.
2. This growth is related to the optimum oxygen tension and is known as a Dinger's ring or disk.⁶

3. Result and Discussion

Leptospirosis, is the most common zoonosis in the world caused by pathogenic spirochetes of the genus leptospira.

Fig. 1: Dinger's ring

Fig. 2: Liquid & semisolid EMJH media

It has been recently recognized as a re-emerging infectious disease among animals and humans and has the potential to become even more prevalent with anticipated global warming. *Leptospira* is related to certain socioeconomic conditions or climatic change that favor endemicity in animal vectors and human exposure. It is generally reported from the developing parts of the world, but cases from industrialized countries following international travel to exotic destinations or involvement of recreational activities were also reported. An estimated global incidence is 5.6 to 50/ 100000 cases, having 300,000- 500,000 severe cases occur each year, with case fatality reports of up to 20-30%.

The clinical presentation is difficult to distinguish from dengue, malaria, influenza, and many other diseases characterized by fever, headache, and myalgia. So difficult to diagnose both by the clinic and the laboratory evidences. Therefore, the disease is frequently not recognized and consequently severely neglected.

Leptospirosis were first noted in South Gujarat, in the Chikhli block of the old Valsad district in the year 1994. Since then cases of leptospirosis are continuously reported from Surat, Navsari and Valsad, Tapi districts of South Gujarat, also from borders from Gujarat. The Factors involved in preponderance of South Gujarat are paddy cultivations and sugarcane (water loving crops), work practice by farmer and animal handler, climatic conditions, clay soil structure leads to water logging higher water tables and rodent carriers. Present study will be useful as less number of data is available regarding prevalent circulating strain of leptospirosis and it's further characterization in South Gujarat.

In present study, patients admitted to New Civil Hospital, Surat with fever, chills conjunctival suffusion, headache, myalgia particularly calf muscles, oliguria, jaundice were included. Total 326 patients included in study during the period of September 2011 to October 2013. The study protocol was approved by the Human Research Ethics Committee of the college. A detailed history and the physical examination findings were recorded on a clinical record form.

Blood culture was taken from all suspected cases of Leptospirosis irrespective of stage/ days of illness or commencement of antibiotics. The culture was done by different methods (i) Whole blood (direct) (ii) whole heparinised blood (iii) surface plasma (iv) the deposit from spun plasma (v) blood clot, inoculated in liquid EMJH medium. All cultures were incubated aerobically and protected from sunlight at room temperature (25° to 30° c) for 6 months. The cultures were examined weekly for 3 months and then every 2 to 4 weeks for 3 months by placing one drop of the culture onto a microscopic slide and observing it by dark-field microscopy at $\times 200$ magnification.

Total 326 blood culture samples were taken during the study period. Different method of blood culture of Leptospirosis were done. Which includes whole blood culture, whole heparinized blood, surface plasma, spun plasma and clot culture. All cultures were sub-cultured according to protocol and screened by dark field microscopy. There were 2 isolates of Leptospirosis from whole blood inoculation, but later due to overgrowth by heavy contamination we could not isolate them in pure culture. So, direct whole blood inoculation at the patients bed side was better culture technique than other blood culture technique used in this study.

Table 1: Shows different methods of culture and there results

Cultures	Whole blood	Whole heparinized blood	Surface plasma	Spun plasma	Clot culture
N=326	2	Nil isolation	Nil isolation	Nil isolation	Nil isolation

Table 2: Shows comparison different methods of culture and there results

	Ana. T. Goncalves ⁹ (2010) (n=68)	Pascale Bourhy ¹⁰ (n=29)	Present study (n=326)
Isolation of Leptospira	10	22	2

Different studies has done to see the which method is better for isolation of Leptospirosis. Study done in 2010 by Ana T. Goncalves shows,⁹ Whole blood inoculation directly in to EMJH medium before antibiotic administration. There isolation rate was 15% (10 / 68 cultured sample). While study by Pascale Bourhy¹⁰ in 2010 used heparinized blood for culture during the acute phase of illness. Ten drops (250 ml) and 20–40 drops (500–1000 ml) of plasma from heparinized blood were transferred into two tubes containing 9 ml of EMJH liquid medium. There isolation rate was 76% (22/29 cultured sample) In the present study we have 2 culture isolates by direct whole blood inoculation (bed-side), unfortunately by overgrown contamination we could not isolate them. Which shows whole blood inoculation is better method for Leptospirosis isolation.

The possible factors involved were, study was conducted at tertiary care hospital, so patient presentation was late in many cases as well as commencement of antibiotic was also the probable factor. For future consideration and better isolation sample for culture should be collected at Primary health care level. Which needs further studies on this topic.

4. Conclusion

Leptospirosis is caused by pathogenic members of the genus *Leptospira*. This infection has a worldwide distribution but it has the greatest impact on health in developing countries, where it is often grossly under diagnosed. Making a diagnosis of leptospirosis in resource-limited settings is challenging because of the similarity between clinical features of other common infectious diseases that occur in the tropics.

The culture method, although with a prolonged incubation time and low diagnostic sensitivity, has an important role in the study of outbreaks and global epidemiology, and provides a crucial pool of clinical strains for studies of pathogenesis.

In present study total 326 blood culture samples from suspected cases of Leptospirosis was collected during the study period. 2 isolates were there but could not isolate in pure culture due to overgrowth of contamination.

5. Acknowledgement

We acknowledge all the staffs of department of Microbiology of Government Medical College, Surat for their full support.

6. Conflict of Interest

We declare that we have no conflict of interest.

References

- Goarant C. Leptospirosis: risk factors and management challenges in developing countries. *Res Rep Trop Med*. 2016;7:49–62. doi:10.2147/rrtm.s102543.
- World Health Organization, WHO, International Leptospirosis Society, ILS. Human Leptospirosis: guidance for diagnosis, surveillance and control. World Health Organization; 2003.
- Dittrich S, Rudgard WE, Woods KL, Silisouk J, Phuklia W, Davong V, et al. The Utility of Blood Culture Fluid for the Molecular Diagnosis of Leptospira: A Prospective Evaluation. *Am J Trop Med Hyg*. 2016;94(4):736–40.
- Limmathurotsakul D, Turner EL, Wuthiekanun V, Thaipadungpanit J, Suputtamongkol Y, Chierakul W, et al. Fool's Gold: Why Imperfect Reference Tests Are Undermining the Evaluation of Novel Diagnostics: A Reevaluation of 5 Diagnostic Tests for Leptospirosis. *Clin Infect Dis*. 2012;55(3):322–31. doi:10.1093/cid/cis403.
- Faine S, Adler B, Bolin C, Perolat P. Leptospira. and others, editor;.
- Levett PN. Leptospirosis. *Clin Microbiol Rev*. 2001;14(2):296–326. doi:10.1128/cmr.14.2.296-326.2001.
- Terpstra WJ. Historical perspectives in leptospirosis. *Indian J Med Microbiol*. 2006;24(4):316. doi:10.4103/0255-0857.29407.
- Wuthiekanunvanaporn C, Limmathurotsakuldirek. Optimization of Culture of Leptospira from Humans with Leptospirosis. *Journal Of Clinical Microbiology*. 2007;p. 1363–1365.
- Ana T. First isolation of human Leptospira strains. *Int J Infect Dis*. 2010;14:148–53.
- Pascale B. 2010.

Author biography

Alpa S Patel, Tutor

Summaiya A Mulla, Professor & HOD

Hardik C Patel, Chief Intensivist

Cite this article: Patel AS, Mulla SA, Patel HC. Leptospira culture and isolation from tertiary care hospital, Surat. *Indian J Microbiol Res* 2021;8(1):24-27.